

## 1D NMR Common Acquisition Concepts and Problems

### (1) Acquisition Parameters and Sampling the FID

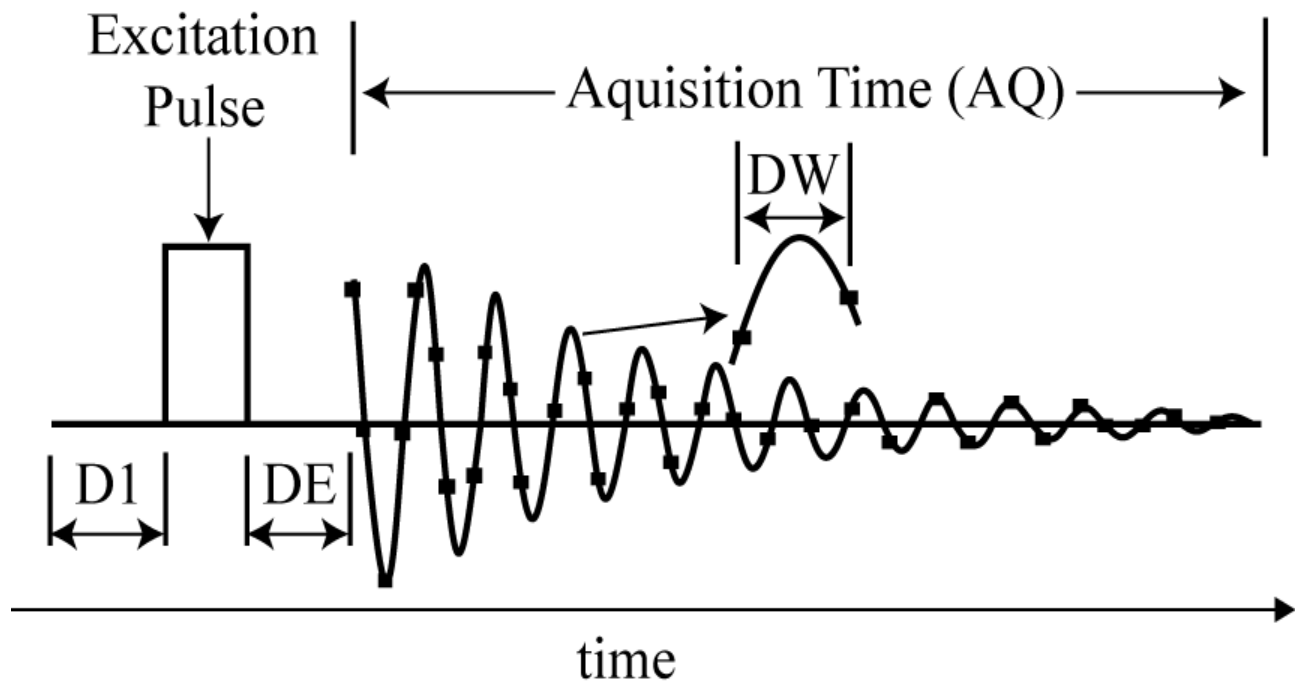
$$\text{Acquisition Time (AQ)} = \text{NumPoints (TD)} * \text{DwellTime (DW)}$$

$$\text{DwellTime (DW)} = 1 / \text{SpectralWidth (SW)}$$

$$\text{Digital Resolutions (FIDRES)} = 1 / [\text{Acquisition Time (AQ)}]$$

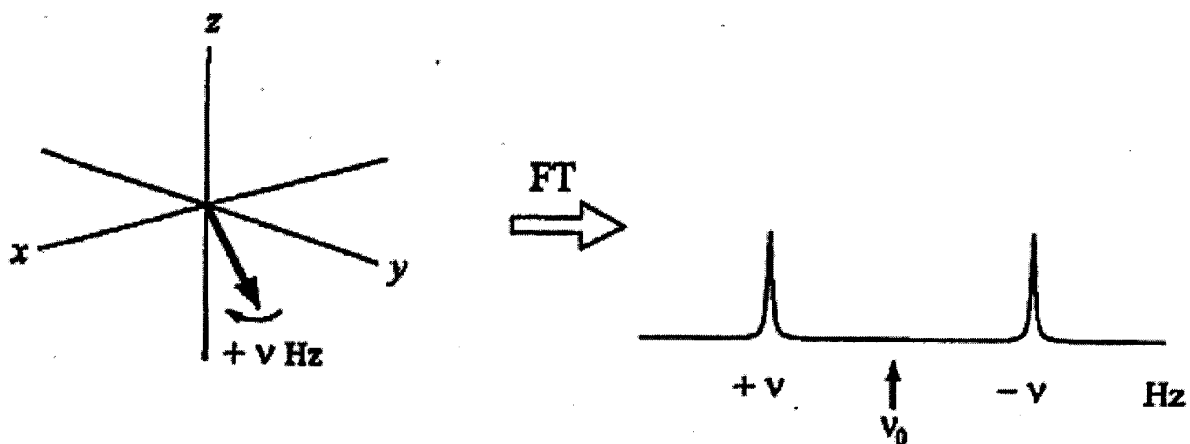
$$\text{D1} = \text{Recycle Delay} \quad \text{DE} = \text{Pre-scan delay}$$

$$\text{NS} = \text{Number of scans} \quad \text{DS} = \text{Number of dummy scans}$$

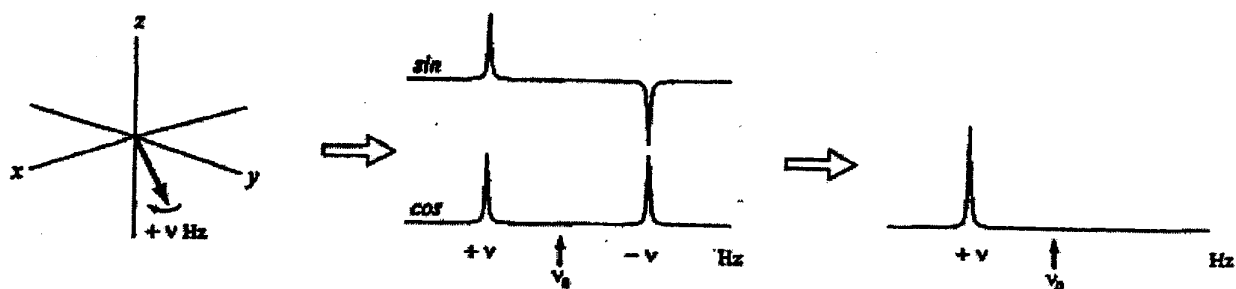


## (2) Quadrature Detection

The spectrum corresponding to a positive precessing magnetization using single channel detection



Detection of a positive rotating magnetization using a two-channel scheme.



### (3) Phasing

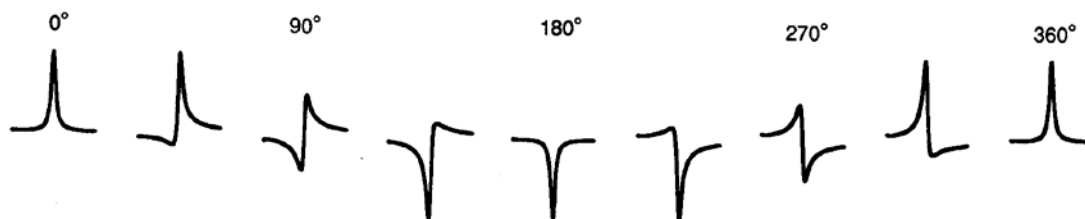
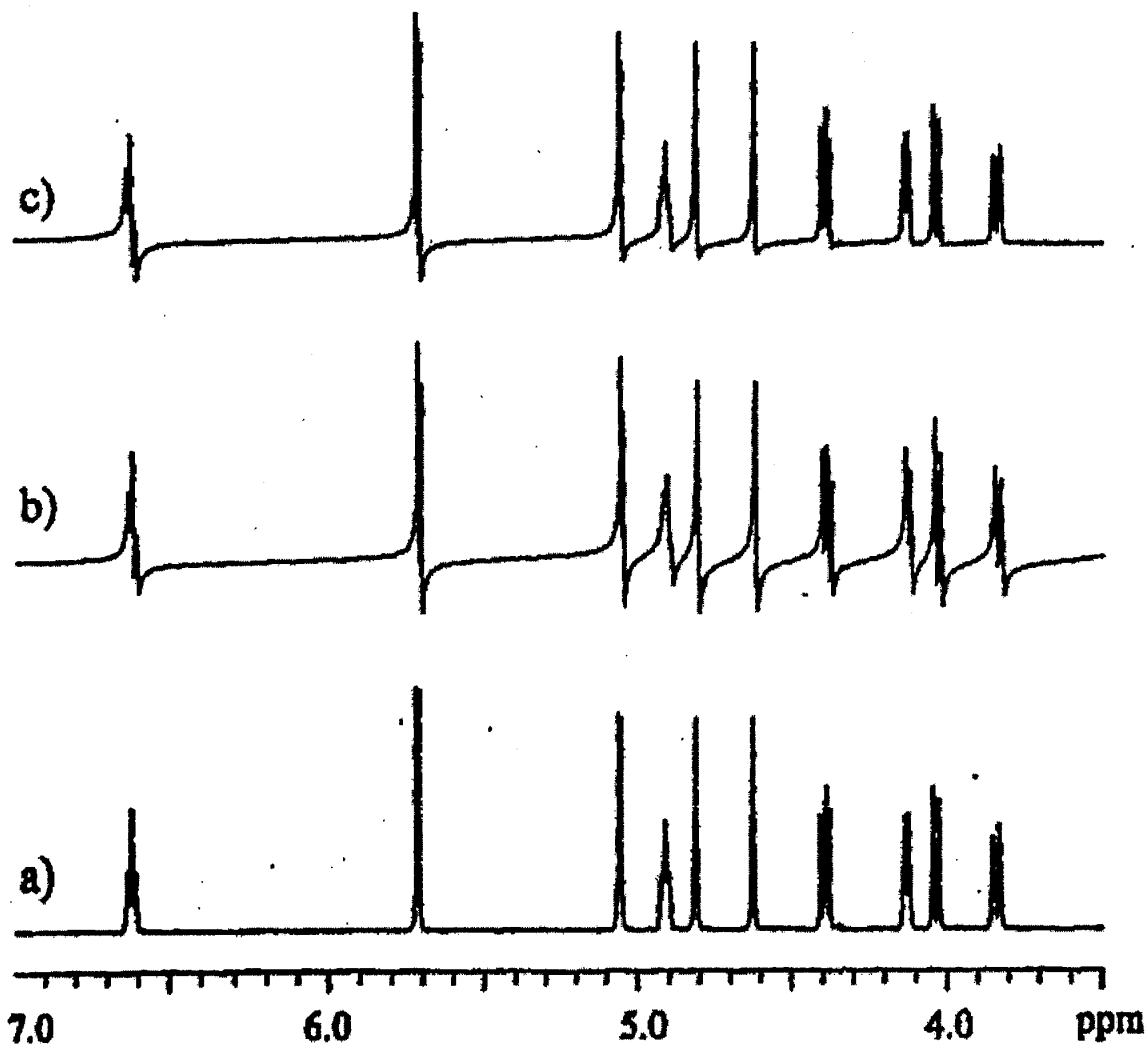
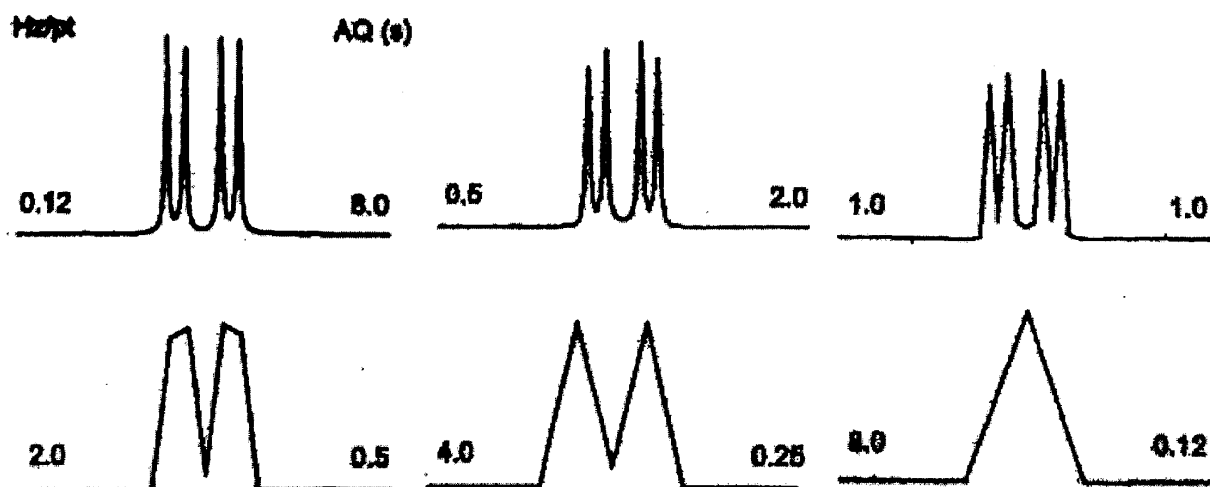


Fig. 1.15. The effect of a change in phase on the shape of a resonance signal; each step in phase is 45°.

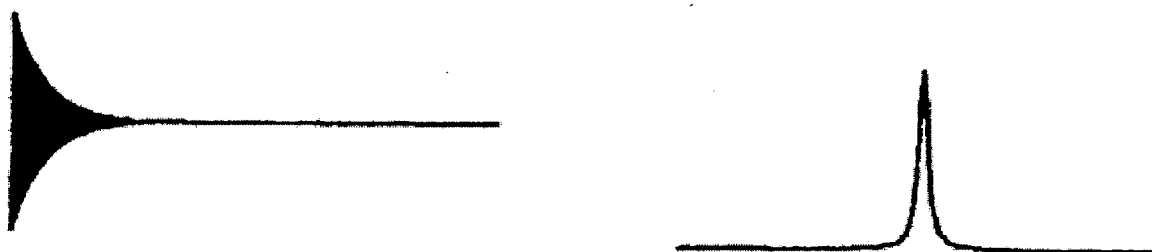
Phase Correcting a Spectrum, a) the initial spectrum, b) after zero-order correction, c) after first-order correction



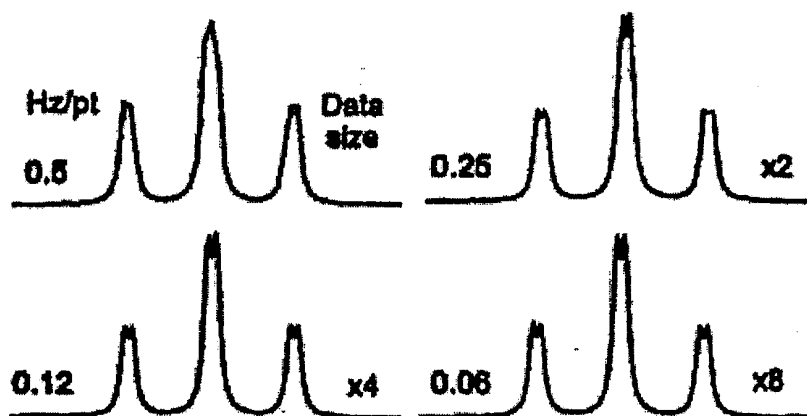
#### (4) Digital resolution



#### (5) Zero filling the FID



Zero filling may reveal fine structure that would otherwise be overlooked



## (6) Exponential Multiplication

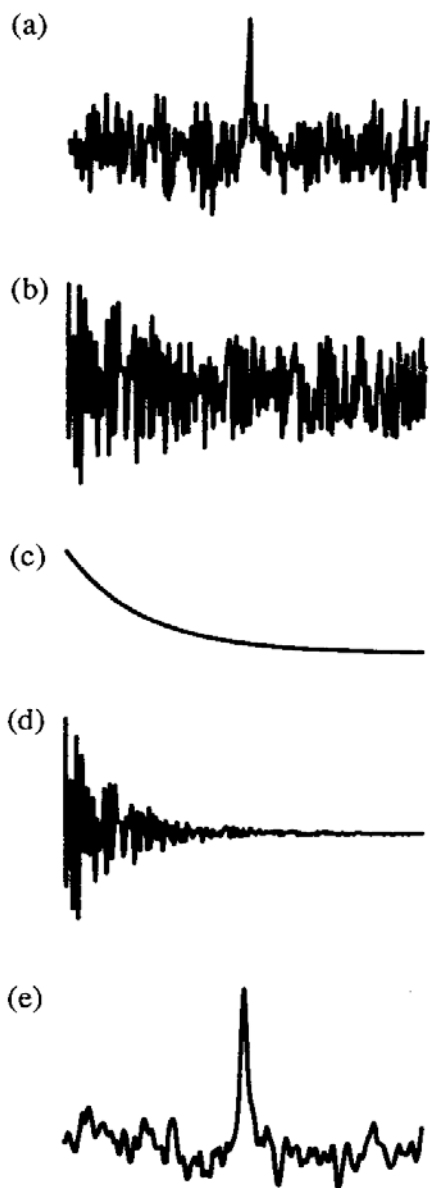


Fig. 1.23. Computer simulation of the effect of exponential multiplication: (a) signal from Fourier transformation of (b); (b) the 'untreated' FID; (c) the exponential decay function; (d) the FID after exponential multiplication; (e) signal from Fourier transformation of (d).

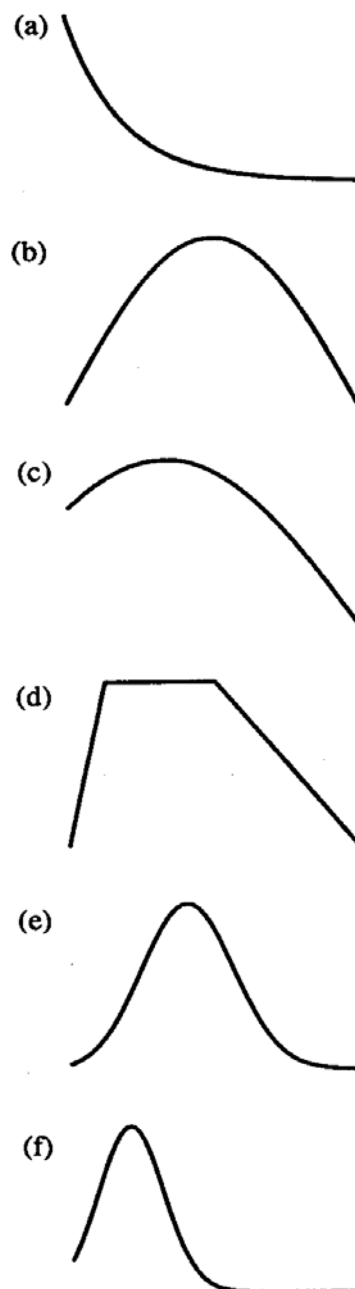
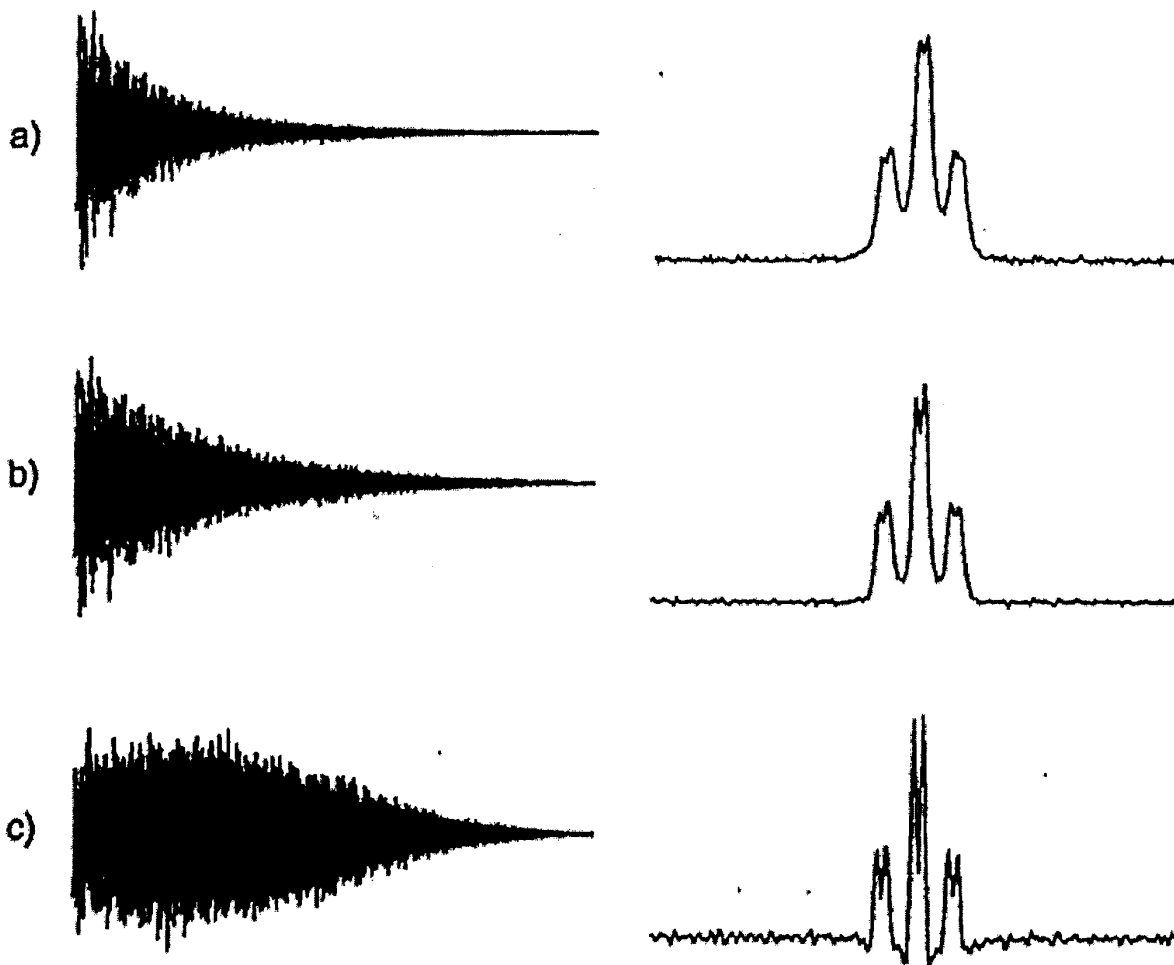


Fig. 1.24. Selected apodization window functions: (a) exponential multiplication; (b) sine-bell; (c) phase-shifted sine-bell; (d) trapezoidal multiplication; (e) Gaussian multiplication; (f) Gaussian multiplication with different parameters.

## (7) Lorentz-Gauss Transformation

- a) Unadulterated FID   b) Exponential Multiplication of FID  
c) Lorentz-Gauss Transformation of FID



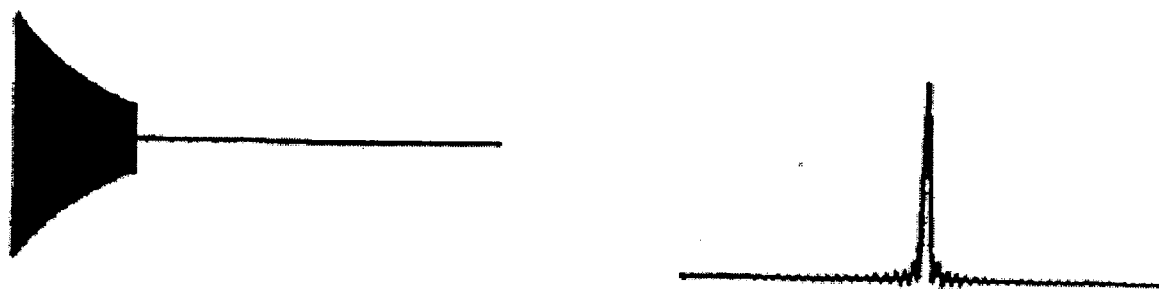
## (8) Folding



Fig. 1.19.  $^1\text{H}$  spectra of furoic acid- $\text{d}_1$ , 1.1. (a) Acquired with the spectral width and reference frequency set correctly; (b) acquired with the reference incorrectly set. The inset shows the 'folded' multiplet.

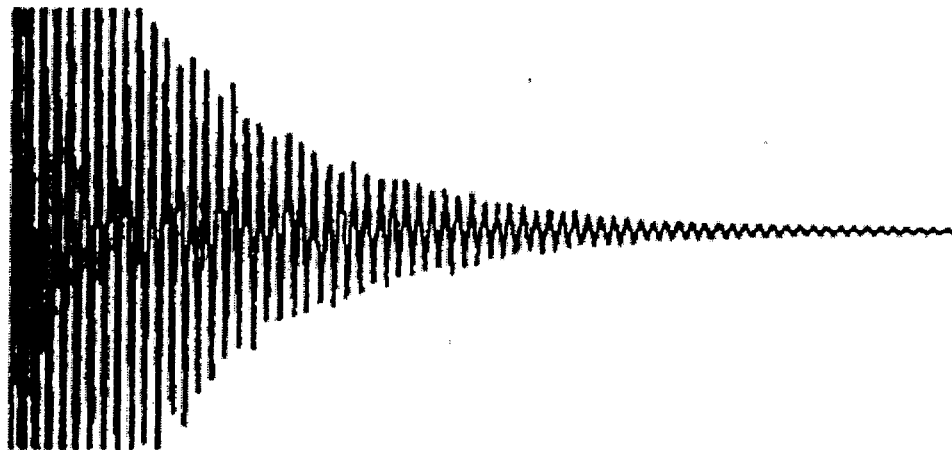
## (9) Truncation

### Truncation with zero filling

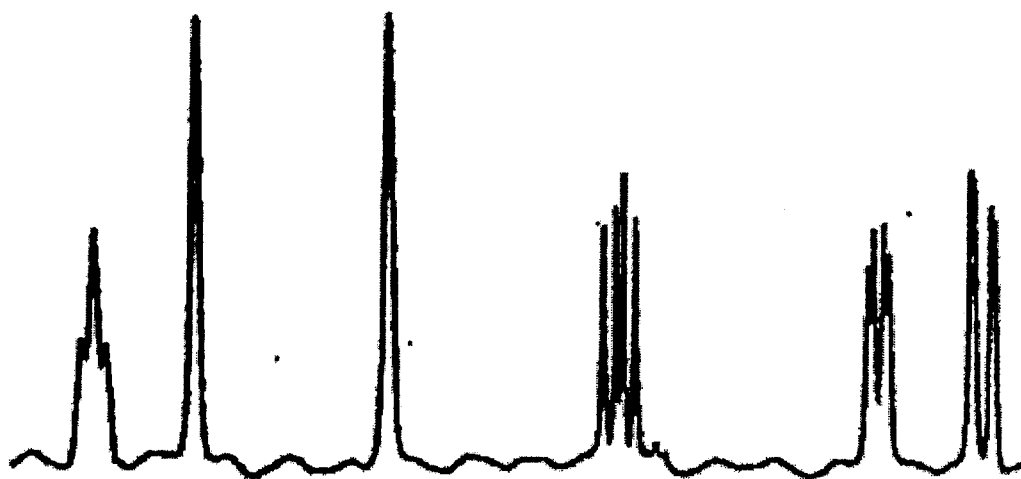


## **(10) Clipping**

**When the signal is too large to digitize the resulting FID is clipped.**



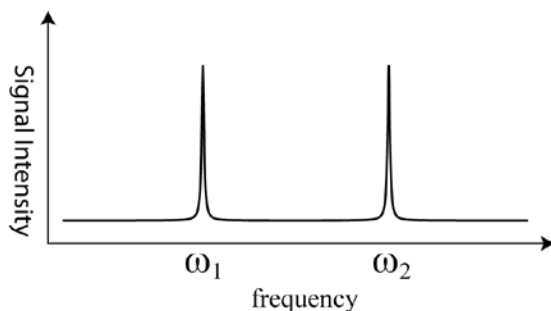
**Baseline Distortions caused by Clipping the FID**





## (11) 2D NMR

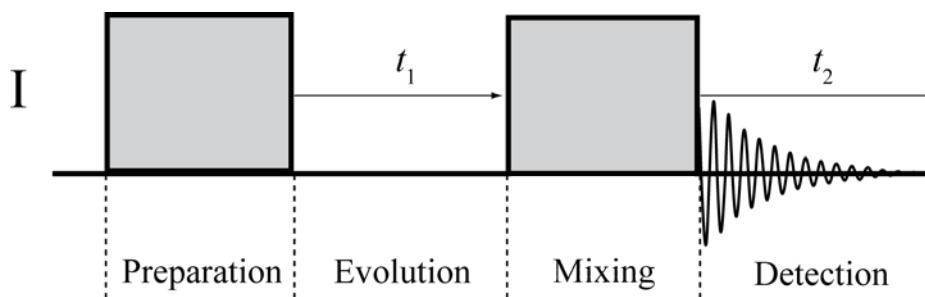
All of the NMR experiments that you have seen up to this point are one-dimensional (1D) techniques where the detected signal is a function of only one time variable. This results in a spectrum that gives the signal intensity as a function of only one frequency axis as seen in figure 29.



**Figure 1A** 1D NMR spectra containing isotropic lines.

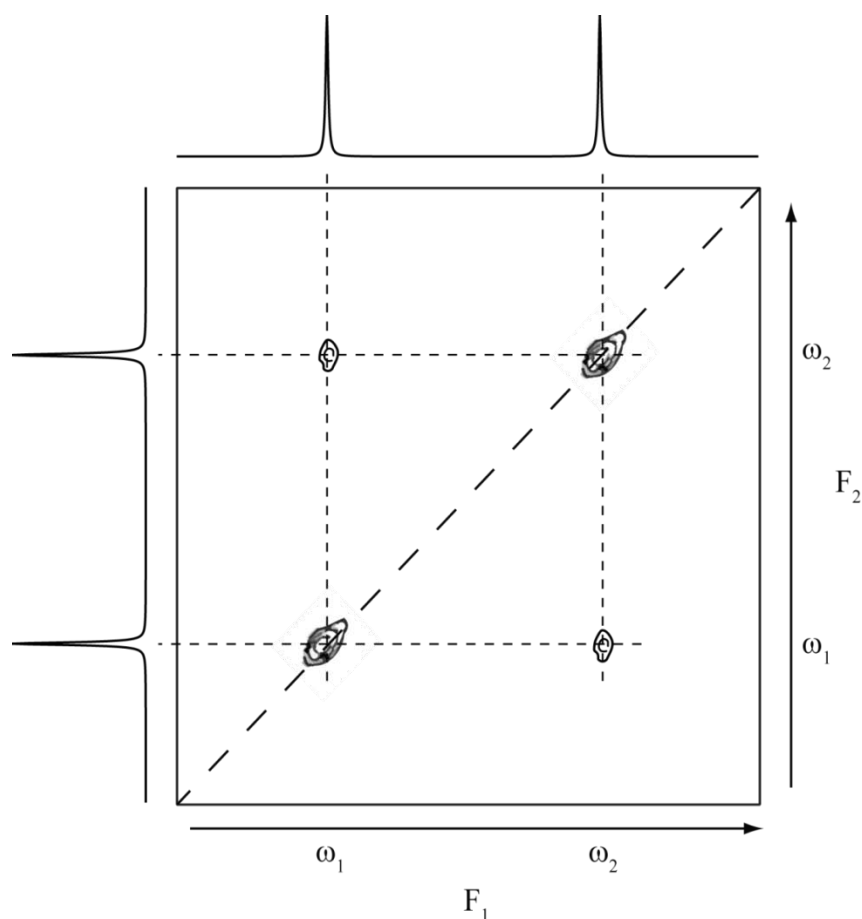
In complex molecules a 1D spectrum can become very crowded, due to multiple chemical shifts, and difficult to interpret; therefore, it is difficult to extract information about the connectivity, or interactions, of the various spin networks. In multi-dimensional NMR spectroscopy the detected signal is a function of several time variables and this provides a tool for analyzing the connectivity of these complex systems.

In two-dimensional (2D) NMR spectroscopy the signal is detected as a function of two time variables, thus providing a way to make observations of the spin-spin interactions directly. The general pulse sequence for a 2D NMR experiment is shown in figure 30 and is composed of four time periods known as preparation, evolution, mixing, and detection.



**Figure 2** The general pulse sequence of a 2D NMR experiment.

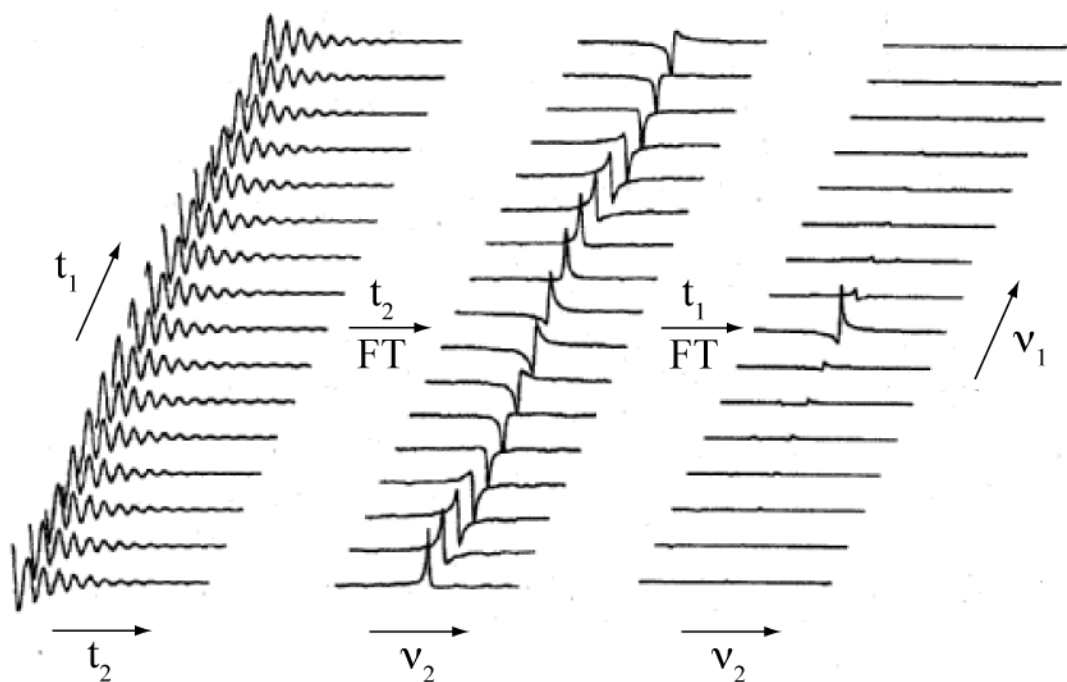
The preparation period uses RF pulses to carefully manipulate the spin system and create a desired state, or coherence order, of the spin system. This state then evolves freely for the duration of the evolution period ( $t_1$ ). It is important to note that the magnetization is not directly detected during the evolution period. The mixing period then uses RF pulses to re-convert the state of the spin system to a form in which it can be detected during the detection period ( $t_2$ ). The resulting FID is now a function of both the evolution ( $t_1$ ) and detection ( $t_2$ ) time periods. The application of a Fourier transform in both time variables results in a 2D spectrum (figure 31) that contains the signal as a function of frequency along two axes,  $F_1$  and  $F_2$ , and these axes correspond to evolution during the time periods of  $t_1$  and  $t_2$ , respectively. The cross-peaks in figure 31 indicate correlations between spins due to coupling.



**Figure 3A** general 2D NMR spectra.

## (12) T1 Evolution

The value of  $t_1$  is incremented, and the sequence is repeated for each point in the indirect time dimension, thereby creating an array of FID's that constitute a data set that is two-dimensional in time  $S(t_1, t_2)$ . This signal is then converted from time domains to the corresponding frequency domains,  $F_1$  and  $F_2$ , via double Fourier transformation, as illustrated in figure 32. Finally, the 2D spectrum is displayed as a contour map plot with frequency axes labeled  $F_1$  and  $F_2$ , correlations between the spins shown as a vertical projection of signal intensities, and the peak coordinates reflecting respective frequencies.



**Figure 32**

Structure of a two-dimensional NMR experiment.